

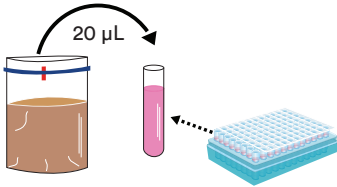
Protocol Reference Guide

Neogen® Molecular Detection Assays 2



1

Invert the capped Lysis Solution (LS) tubes to mix.



2

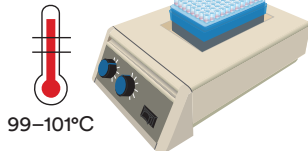
De-cap lysis tube strip and discard rubber cap.
Do not re-cap the LS tubes.

Transfer 20 µL sample to individual LS tube.
If lysate will be retained for re-test, place caps into a clean container for later use.



Liquid will be pink.

00:15:00



3

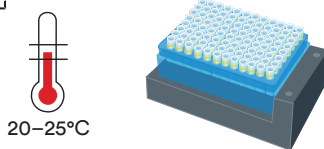
Heat uncovered LS tube rack on heating block at 100°C (±1°C) for 15 minutes.

Do not re-cover the LS tube rack.



Liquid will be yellow during heating.

00:05:00



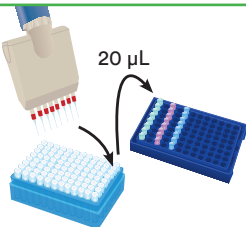
4

Cool LS tube rack without lid on chill block at room temperature (20–25°C) for 5 minutes.

Do not freeze the chill block nor use the tray.

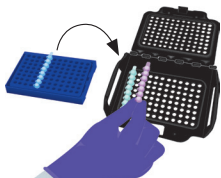


Liquid will be pink when cooled.



5

Transfer 20 µL sample to individual reagent tube.
Pipet up and down 5 times to mix.



6

Transfer closed tubes to speed loader tray.

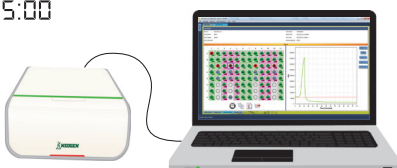


7

Start the run using the software. Place speed loader tray into instrument and close the lid.

If lysate will be retained for re-test, wash hands, re-glove and then re-cap the LS tubes after this step. Store capped LS tubes at 4–8°C.

00:75:00



8

Presumptive positive results as early as 15 minutes.
Negative results at 60–75 minutes.

Salmonella, E. coli O157 (including H7), Cronobacter, Campylobacter, STEC Gene Screen (stx) and STEC Gene Screen (stx and eae) at 60 minutes.
Listeria and Listeria monocytogenes at 75 minutes.